

MagIso™ Anti-MERS-CoV Spike Protein S2 Magnetic Particles Immunoprecipitation (IP) Kit

Catalog Number: WHK-SN014

Please read this instruction manual carefully before using the product

Product Contents

	Contents	Package 1 (20 Tests)	Package 2 (100 Tests)	Storage
1	Anti-MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 Magnetic Beads ^{1,3}	1 mL	5 mL	2-8°C for 12 months
2	NP40 Cell Lysis Buffer ²	4 mL	22 mL	-20°C for 12 months
3	5×TBST (pH7.4)	Required but not supplied		
4	1×TBST (pH7.4)	Required but not supplied		
5	ddH ₂ O	Required but not supplied		
6	Alkaline Elution Buffer	3 mL	15 mL	2-8°C for 12 months
7	Acidity Elution Buffer	3 mL	15 mL	2-8°C for 12 months
8	Neutralization Buffer	2 mL	8 mL	2-8°C for 12 months

[1] The IP KIT contains anti-MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 Immunomagnetic Beads(2 mg/mL) in phosphate buffered saline (PBS, pH 7.4) with sodium azide(0.1%).

[2] Using NP-40 cell lysate buffer in the kit is required, otherwise, the magnetic beads may be precipitated.

[3] Immunomagnetic Beads kits are shipped at ambient temperature in which immunomagnetic beads are provided in liquid buffer.

Product Description

The Anti-MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 Immunomagnetic Beads, conjugated with Anti-MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 antibody, are used for immunoprecipitation (IP) of MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 proteins which expressed in vitro expression systems and bacterial and mammalian cell lysates.

For IP, the beads are added to a sample containing MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 proteins to form a bead-protein complex. The complex is removed from the solution manually using a Magnetic Separator. The bound MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 proteins are dissociated from the Immunomagnetic Beads using an elution buffer.

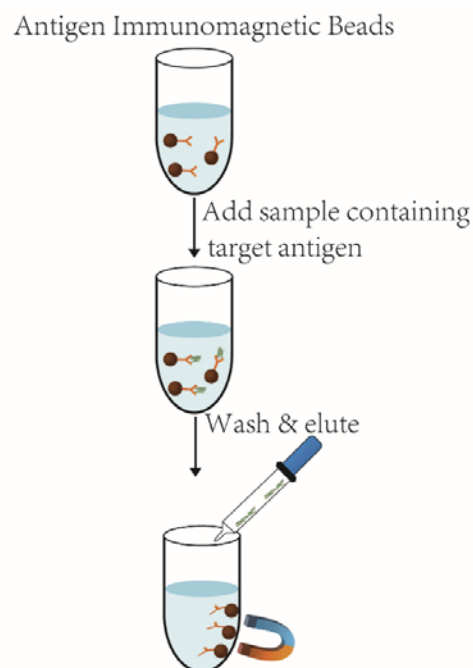


Fig. 1 Immunoprecipitation (IP) Protocol

Antibody Information

Antibody: MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 Antibody, Mouse MAb

Immunogen: Recombinant MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 Protein

Clone ID: 11

Isotype: Mouse IgG1

Specificity: MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2

Preparation: This antibody was produced from a hybridoma resulting from the fusion of a mouse myeloma with B cells obtained from a mouse immunized with purified, recombinant MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2. The IgG fraction of the cell culture supernatant was purified by Protein A affinity chromatography.

Applications: IP, Minimum Protein Purification

Notes: The applications have not been validated with corresponding viruses.

Protocol

The protocol (Fig. 1) uses 50 μ L Anti-MERS-CoV (NCoV / Novel coronavirus) Spike Protein S2 Immunomagnetic Beads, but this can be scaled up or down as required.

Cell Lysis

Cells may be lysed using any standard cell lysis protocol in accordance with your starting materials. **We suggest using NP40 Cell Lysis Buffer (supplied with kit).**

Immunoprecipitate Target Antigen

1. Add 50 μ L of Immunomagnetic Beads into a 1.5 mL microcentrifuge tube.
2. Add 150 μ L of 1 \times TBST buffer to the Immunomagnetic Beads and gently vortex to mix.
3. Place the tube into a Magnetic Separator to collect the beads against the wall side of the tube. Remove and discard the supernatant.
4. Add 1 mL of 1 \times TBST buffer to the tube. Invert the tube several times or gently vortex to mix for 1 min. Collect Immunomagnetic Beads with a Magnetic Separator. Remove and discard the supernatant.
5. Add the sample containing target protein (Cell lysate: 0.5-1mg; Recombinant protein: 5-25 μ g) to the pre-washed Immunomagnetic Beads, add 1 \times TBST buffer until final

volume to 200-500 μ L, and incubate at 37°C for 20-30 min (or at room temperature for 2-3h) with mixing.

5. Collect the Immunomagnetic Beads with a Magnetic Separator, remove the unbound sample and save for analysis.

6. Add 300 μ L of 5 \times TBST buffer to the tube and gently mix. Collect the Immunomagnetic Beads and discard the supernatant. Repeat this wash twice.

7. Add 300 μ L of ddH₂O to the tube and gently mix. Collect the Immunomagnetic Beads on a Magnetic Separator and discard the supernatant.

Elute Target Antigen.

A. Alkaline Elution

1. Add 100 μ L of Alkaline Elution buffer to the tube.
2. Gently vortex to mix and incubate the sample at room temperature on a rotator for 5 min.
3. Magnetically separate the Immunomagnetic Beads and save the supernatant containing the target antigen.
4. To neutralize the sample, add 50 μ L of Neutralization Buffer for each 100 μ L of eluate.

B. Acidity Elution

1. Add 100 μ L Acidity Elution Buffer.
2. Gently vortex to mix and incubate the sample at room temperature on a rotator for 5-10 min.
3. Magnetically separate the Immunomagnetic Beads and save the supernatant containing the target antigen.
4. To neutralize the low pH, add 15 μ L of Neutralization Buffer for each 100 μ L of eluate.

C. Denaturing Elution

1. Add 10 μ L of 2 \times SDS-PAGE Sample Loading Buffer to the tube.
2. Gently vortex to mix and incubate the sample at 95-100 °C for 5-10 min.
3. Magnetically separate the Immunomagnetic Beads and save the supernatant containing the antigen.

General Test System (for reference):

	Recombinant Protein	Cell Lysate
Sample Quality	10 μ g add into 0.5mg cell lysate (without interfering proteins)	0.5mg
Final Volume	300 μ L	
Incubate Time	Room temperature, 2h	
Elute	Using 10 μ L of 2 \times SDS-PAGE Sample Loading Buffer	

Trouble Shooting

Problem	Possible Cause	Solution
Magnetic Beads aggregated	Magnetic Beads were frozen or centrifuged	Handle the Beads as directed in the instructions
	Buffer was incompatible with magnetic beads	
	Detergent was not added to the wash and bind solutions	
Failure to co-IP interacting protein	Wash conditions were too stringent for the weak or transient interaction	Reduce the number of washes
		Lower the ionic strength of the wash buffer
	Interacting protein was expressed at a low level	Apply additional protein sample
		Use a more sensitive detection system
	Buffer system was not optimal for the protein: protein interaction	Optimize the co-IP buffer
	Insufficient sample was loaded on the gel for Western blot detection	Elute sample in 30% acetonitrile 0.5% formic acid, then
Bring the sample back up in SDS-PAGE Sample Loading Buffer and load entire elution fraction on		

Problem	Possible Cause	Solution
Little or no protein is detected	Protein degraded	Include protease inhibitors (PMSF) in the lysis buffer
		Use new lysate or lysate stored at -80° C
	No or minimal protein was expressed	Verify protein expression by SDS-PAGE or Western blot
		Analysis of the lysate using an positive control as a reference
		Increase the amount of lysate used for IP/Co-IP Use a more sensitive detection system